



Genome-wide association study of brain amyloid deposition as measured by Pittsburgh Compound-B (PiB)-PET imaging

Qi Yan, PhD^{1,2}, Kwangsik Nho, PhD^{3,4}, Jorge L. Del-Aguila, PhD⁵, Xingbin Wang, PhD¹, Shannon L. Risacher, PhD^{3,4}, Kang-Hsien Fan, PhD¹, Beth E. Snitz, PhD^{6,7}, Howard J. Aizenstein, PhD⁸, Chester A. Mathis, PhD^{7,9}, Oscar L. Lopez, MD^{6,7,8}, F. Yesim Demirci, MD¹, Eleanor Feingold, PhD¹, William E. Klunk, MD, PhD^{6,7,8}, Andrew J. Saykin^{3,4} on behalf of Alzheimer's Disease Neuroimaging Initiative (ADNI), Carlos Cruchaga, PhD⁵, and M. Ilyas Kamboh, PhD^{1,7,8}

¹Department of Human Genetics, University of Pittsburgh, Pittsburgh, PA USA

²Department of Pediatrics, Children's Hospital of Pittsburgh of UPMC, University of Pittsburgh, Pittsburgh, PA USA

³Department of Radiology and Imaging Sciences, Indiana University School of Medicine, Indianapolis, IN USA

⁴Indiana Alzheimer Disease Center, Indiana University School of Medicine, Indianapolis, IN USA

⁵Department of Psychiatry, Washington University School of Medicine, St. Louis, MO USA

⁶Department of Neurology, University of Pittsburgh, Pittsburgh, PA USA

⁷Alzheimer Disease Research Center, University of Pittsburgh, Pittsburgh, PA USA

⁸Department of Psychiatry, University of Pittsburgh, Pittsburgh, PA USA

⁹Department of Radiology, University of Pittsburgh, Pittsburgh, PA USA

Abstract

Deposition of amyloid plaques in the brain is one of the two main pathological hallmarks of Alzheimer's disease (AD). Amyloid positron emission tomography (PET) is a neuroimaging tool that selectively detects *in vivo* amyloid deposition in the brain and is a reliable endophenotype for AD that complements cerebrospinal fluid biomarkers with regional information. We measured *in vivo* amyloid deposition in the brains of ~1,000 subjects from three collaborative AD centers and ADNI using ¹¹C-labeled Pittsburgh Compound-B (PiB)-PET imaging followed by meta-analysis of genome-wide association studies, first to our knowledge for PiB-PET, to identify novel genetic

Users may view, print, copy, and download text and data-mine the content in such documents, for the purposes of academic research, subject always to the full Conditions of use:http://www.nature.com/authors/editorial_policies/license.html#terms

Address for correspondence: M. Ilyas Kamboh, PhD, Department of Human Genetics, Graduate School of Public Health, University of Pittsburgh, PA 15261, USA, kamboh@pitt.edu.

CONFLICT OF INTEREST

GE Healthcare holds a license agreement with the University of Pittsburgh based on the PiB-PET technology described in this manuscript. Drs. Klunk and Mathis are co-inventors of PiB and, as such, have a financial interest in this license agreement. GE Healthcare provided no grant support for this study and had no role in the design or interpretation of results or preparation of this manuscript. All other authors have no conflicts of interest.

loci for this endophenotype. The *APOE* region showed the most significant association where several SNPs surpassed the genome-wide significant threshold with *APOE*4* most significant (P -meta=9.09E-30; β =0.18). Interestingly, after conditioning on *APOE*4*, 14 SNPs remained significant at $P<0.05$ in the *APOE* region that were not in linkage disequilibrium with *APOE*4*. Outside the *APOE* region, the meta-analysis revealed 15 non-*APOE* loci with $P<1E-05$ on nine chromosomes, with two most significant SNPs on chromosomes 8 (P -meta=4.87E-07) and 3 (P -meta=9.69E-07). Functional analyses of these SNPs indicate their potential relevance with AD pathogenesis. Top 15 non-*APOE* SNPs along with *APOE*4* explained 25%–35% of the amyloid variance in different datasets; of which 14–17% was explained by *APOE*4* alone. In conclusion, we have identified novel signals in *APOE* and non-*APOE* regions that affect amyloid deposition in the brain. Our data also highlights the presence of yet to be discovered variants that may be responsible for the unexplained genetic variance of amyloid deposition.

Keywords

Brain amyloid; amyloid-PET; GWAS; meta-analysis

INTRODUCTION

Genomic efforts mainly through large-scale genome-wide association studies (GWAS), as part of the Alzheimer's Disease Genetics Consortium (ADGC)¹ and the International Genomics of Alzheimer's Project (IGAP)² have identified over 20 genes/loci for late-onset Alzheimer's disease (AD). However, known common AD variants account for only ~30% of the AD genetic variance³ and they also do not provide definitive information about underlying disease mechanisms. Genetic studies focusing on AD-related quantitative phenotypes/endophenotypes may help to identify additional AD-related genes. One such AD-related phenotype is deposition of amyloid-beta ($A\beta$) in the brain, which is one of the two main pathologic hallmarks of AD; the other being the formation of tau deposits in the form of neurofibrillary tangles, neuropil threads and dystrophic neurites (tau pathology) in the brain.⁴ According to the current model for sporadic AD, $A\beta$ pathology occurs independently of tau pathology, is detectable earlier and is believed to accelerate neocortical tau pathology and neurodegeneration.⁵ Recent longitudinal studies on cognitively normal subjects also confirm that amyloidosis is an early process in AD.^{6, 7} The *in vivo* detection of $A\beta$ deposition in the brain, as measured by positron emission tomography (PET) scanning with ¹¹C-labeled Pittsburgh Compound-B (PiB) and the increased retention of PiB observed in the brains of AD patients compared to cognitively normal controls, was first reported by Klunk and colleagues^{8, 9} and since has been confirmed in many studies.¹⁰ There is a high correlation between amyloid PET imaging and neuritic plaque frequency as confirmed by autopsy studies.^{11–13} Multiple studies have shown that amyloid PET has a high value for the clinical diagnosis of AD and in clinical trials aiming to reduce brain $A\beta$ burden.¹⁴

There is a well-established association of *APOE* variants with risk^{1, 2} and age-at-onset^{15, 16} of AD. Likewise, *APOE* genetic variation is also strongly associated with $A\beta$ deposition in the brain as measured by PiB retention,^{17–19} indicating a genetic basis of $A\beta$ deposition in the brain. Here, we used PiB-PET as an endophenotype to identify novel genetic loci for AD

pathology using meta-analysis of three GWAS, the first to our knowledge, using the largest sample with the PiB-PET imaging from three different centers and the Alzheimer's Disease Neuroimaging Initiative (ADNI).

MATERIALS AND METHODS

Sample description

All subjects with PiB-PET data were European-Americans and derived from three sites: University of Pittsburgh (PITT), Washington University (WU) and Indiana University (IU) combined with the initial phase of the multicenter ADNI PiB-PET add-on study (here they are referred to as ADNI/IU). All subjects provided informed consent and all studies were approved by their local Institutional Review Boards. The summary statistics of these samples are included in Supplementary Table S1 and their description is given Supplementary Text.

Amyloid-PET data

Detailed methods for acquisition and processing of PiB-PET scans are described in previous reports for the PITT,^{17, 18} WU,¹⁹ ADNI²⁰⁻²² and IU²³ studies. PiB retention was measured in 4 cortical regions of the brain, including medial frontal cortex (MFC; anterior cingulate/gyrus rectus), lateral frontal cortex (LFC), precuneus cortex (PRC) and parietal cortex (PAR) and expressed as a ratio to the cerebellum. In the GWAS meta-analysis, the PiB retention values from these 4 cortical regions were averaged in each subject to calculate a mean global score (GBL4) as the quantitative endophenotype. PiB retention was expressed as standardized uptake volume ratio (SUVR) in the PITT and ADNI/IU data^{23, 24} and as binding potential (BP) in the WU data.²⁵ BP is approximately equal to SUVR-1. Because of this inconsistency in the PiB measurement methods the GWAS data were analyzed via *P*-value based meta-analysis as described below.

Genotyping, imputation and quality control

The genotyping platforms used for each study sample are listed in Supplementary Table S1. Imputation of non-genotyped single-nucleotide polymorphisms (SNPs) was performed with IMPUTE2²⁶ using the 1000 Genomes Project²⁷ Phase III (May 2013 release) data as the reference panel for PITT and Phase I (November 2010 release) data for WU and ADNI/IU datasets. Full description of these procedures is given in Supplementary Text.

Meta-analysis

METAL²⁸ software was used to perform meta-analysis on three GWAS using the mean PiB-PET GBL4 value. METAL performs a *P*-value based meta-analysis, which is appropriate when the effects being estimated are different in different cohorts. It does, however, account for differences in sample size between cohorts and for the direction of effects. The summary effect size was calculated by averaging the study-specific effect sizes, with weights reflecting the standard errors from the study-specific effect sizes.

Functional analyses

To evaluate the biological significance of PiB-associated signals, we conducted five different analyses: differential gene expression in AD versus non-AD in relevant tissues, brain gene expression, expression quantitative trait loci (eQTL) analyses, summary-data-based Mendelian randomization (SMR) analyses to test for pleiotropic association between gene expression and PiB, and pathway analyses. Detailed description of these analyses is given in Supplementary Text.

RESULTS

Amyloid PET data characteristics

The characteristics of participants in each of the three datasets included in the meta-analysis are shown in Supplementary Table S1. The WU sample was younger with less male participants. The distribution of mean global PiB retention is shown in Figure 1.

GWAS Analysis

Quantile-quantile (QQ) plots and lambda values for the meta-analysis showed that neither the results from each of the three component studies nor the combined results from meta-analysis were inflated in their test statistics (Figure 2a). Meta-analysis revealed 27 genome-wide significant SNPs ($P < 5E-08$) in a four-gene region on chromosome 19: *PVRL2-TOMM40-APOE-APOC1* (Figure 2b, and Supplementary Table S2). As expected, *APOE*4*/rs429358 showed the most significant association with the average global PiB retention ($P_{\text{meta}} = 9.09E-30$; $\beta = 0.18$; Figure 3, Supplementary Figure S1).

Outside of the *APOE* region, no genome-wide significant signal was observed. However, the meta-analysis revealed 15 non-*APOE* loci with $P < 1E-05$ on chromosomes 8, 3, 15, 4, 21, 13, 2, 12 and 1 (Table 1). Most of these loci show quite consistent results across the all datasets. The regional plots of these 15 non-*APOE* loci are shown in Supplementary Figures S2.1-S2.15. The most significant SNP outside the *APOE* region is intergenic located between *ADCY8 - EFR3A* on chromosome 8 (rs13260032; $P = 4.87E-07$, Supplementary Figure S2.1). The next most significant SNP is also intergenic located between *RAP2B - C3orf79* on chromosome 3 (rs4680057; $P = 9.69E-07$, Supplementary Figure S2.2). Chromosome 3 also harbors two additional signals: one in ncRNA (*LINC00971*/rs9831119; $P = 2.98E-06$, Supplementary Figure S2.6) and another near *MAGEF1*/rs11923588 ($P = 5.66E-06$, Figure S2.9). The third most significant SNP is located in the *DAPK2* gene on chromosome 15 (rs12908891; $P = 1.39E-06$, Supplementary Figure S2.3). We also analyzed the data after adjusting for the effect of *APOE*4*/rs429358 in these non-*APOE* regions, which showed a slight attenuation of the association strengths (Table 1).

Conditional Analysis in the *APOE* Region

In order to check if there were independent SNPs associated with the PiB retention in the *APOE* region, we performed conditional analysis by adjusting for the top SNP (*APOE*4*/rs429358). A total of 14 SNPs remained significant at $P < 0.05$ (Table 2), including 3 SNPs that showed genome-wide significance before adjusting for *APOE*4* (rs75627662, rs483082 and rs438811; Supplementary Table S2). Supplementary Figure S3 shows LD structure of

these 14 SNPs along with *APOE**4/rs429358 and *APOE**2/rs7412 SNPs. *APOE**4 and *APOE**2 have essentially no LD with 9 of the 14 SNPs that are located in the *PVRL2* gene (SNPs 1–9 in Supplementary Figure S3). One SNP located in the *APOE/APOC1* intergenic region (rs59325138) has only very weak correlation with *APOE**4 ($R^2=0.15$) and *APOE**2 ($R^2=0.03$). While 3 SNPs located down stream of *APOE* and *APOE/APOC1* intergenic region have weak to moderate LD with *APOE**4 ($R^2=0.42, 0.64, 0.65$ for rs75627662, rs483082 and rs438811, respectively).

The most significant SNP in meta-conditional analysis was *APOE**2/rs7412 (P -meta=3.69E-03; $\beta=-0.06$; Table 2), though it was not genome-wide significant before adjusting for *APOE**4 (P -meta=6.57E-05; $\beta=-0.09$). A similar strength of association was seen with an intronic *PVRL2*/rs3852859 SNP after adjusting for *APOE**4 (P -meta=8.8E-03; $\beta=0.06$; Table 2) that was in LD with 3 additional SNPs (SNPs 1, 7, 9 in Supplementary Figure S3). Three additional apparently independent associations were seen with rs4803767 (P -meta=2.06E-02; $\beta=0.05$ Table 2) that was in LD with 4 additional SNPs (SNPs 2–5 in Supplementary Figure S3), rs75627662 (P -meta=1.50E-02; $\beta=-0.03$; Table 2) that was in LD with 2 additional SNPs (SNPs 13,15 in Supplementary Figure S3) and rs59325138 (P -meta=3.10E-02; $\beta=0.03$; Table 2) that has very weak correlation with all other SNPs ($R^2=0.01-0.24$).

Association of known AD risk loci with amyloid burden and Association of amyloid loci with AD risk

We examined the top IGAP genome-wide significant SNPs (Supplementary Table S3.1) and the associated gene regions (Supplementary Table S3.2) in relation to amyloid burden and found only some nominally significance SNPs. Likewise, we examined the suggestive non-*APOE* amyloid loci in our PITT-ADRC case-control sample of >2,200 subjects²⁹ and found association of two top amyloid-associated SNPs with AD risk (Supplementary Table S4.1). When we examined additional A β -associated SNPs in each region with AD risk, we found multiple associations with $P<0.05$ (Supplementary Table S4.2), indicating that our suggestive A β -associated loci are also associated with AD risk (see Supplementary Text for more details).

Estimation of Amyloid-PET variance by *APOE* and non-*APOE* loci

The genetic variance was estimated based on the R-square calculated from a linear regression model regressing global PiB retention on 6 independent *APOE* SNPs (rs429358, rs7412, rs3852859, rs4803767, rs75627662, rs59325138), as described above, and 15 non-*APOE* SNPs given in Table 1. The contribution of 6 *APOE* SNPs to the variance of global PiB retention was 28.0%, 17.3% and 17.12% in the PITT, WU and ADNI/IU datasets, respectively; *APOE**4/rs429358 alone explained 17.5%, 16.5% and 13.9%, respectively. The top 15 non-*APOE* SNPs explained 22.6%, 21.6% and 21.7% of the amyloid variance in the PITT, WU and ADNI/IU datasets, respectively. The consistency of these estimates across the different datasets gives confidence that the difference in measurement of PiB across the datasets does not affect the bottom-line results.

Functional analyses

We performed five analyses (see Methods section) to evaluate the biological significance of PiB-implicated signals/genes. We considered all genes within $\pm 500\text{kb}$ of the top variant in each locus from Table 1 plus any eQTL-controlled genes outside the $\pm 500\text{kb}$ boundary as target genes (Figure 3, Supplementary Figures S2.1-S2.15), and selected a total of 257 genes.

Of 257 target genes, we found 20 upregulated and 25 downregulated genes that were differentially expressed in the same direction in two or more AD studies and no opposite directions were reported (Table 3 and Supplementary Table S6 marked in green color). Brain RNA-seq data reveals many of these differentially expressed candidate genes are expressed in AD-relevant cell types (Table 3 and Supplementary Table S6 marked in yellow color).

For eQTL analyses, we identified SNPs in LD ($R^2 \geq 0.5$) with the top SNP for each locus in Table 1. For these SNPs, there were *cis*-acting eQTLs (eQTL $P < 0.05$) for 151 of the 257 target genes in various brain tissues, and 36 genes in whole blood available in GTEx. Supplementary Table S5 gives the eQTL results for each top SNP in 15 non-*APOE* loci and the detailed results of LD SNPs ($R^2 \geq 0.80$) with top SNPs are given in Supplementary Table S7. With the exception of *SLITRK1* /rs9831119, the other 14 top SNPs were eQTLs in different brain regions; 11 of them were eQTL in anterior cingulate cortex/frontal cortex/cortex where PiB intake is highest,³⁰ indicating their role in affecting amyloid deposition in the brain.

For SMR analyses, only the gene/variant pairs identified in the *cis*-eQTL analyses were considered. For these gene/variant pairs, 99 genes in any brain tissue and 19 in whole blood were shown to mediate genetic effects on PiB by *cis*-regulating gene expression (SMR $P < 0.05$; Table 3, Supplementary Table S6).

We conducted pathway analyses (MAGMA³¹) using four gene set resources, including and excluding target genes in the *APOE* region, and detected nine genome-wide significant pathways: ndkdynammin pathway, $FDR = 4.6E-04$; synaptic vesicle recycling, $FDR = 3.5E-07$; synaptic vesicle endocytosis, $FDR = 3.1E-04$; protein depolymerization, $FDR = 3.1E-04$; inositol tetrakisphosphate phosphatase activity, $FDR = 5.7E-03$; positive regulation of vacuole organization, $FDR = 5.7E-03$; inositol trisphosphate phosphatase activity, $FDR = 0.033$; regulation of clathrin-mediated endocytosis, $FDR = 0.038$; and clathrin-mediated endocytosis, $FDR = 0.043$. Although none of the 257 target genes, including *APOE*, are included in these nine genome-wide significant pathways, 71 target genes are included in the nominally significant pathways and 46 target genes are included in the non-*APOE* region-related nominally significant pathways ($P < 0.05$. Table 3 and Supplementary Table S6 marked in pink color).

DISCUSSION

In this investigation, we have used the largest PiB-PET imaging data ($n \sim 1,000$), available from multiple collaborative centers, as an endophenotype to identify novel genetic loci for

AD pathology using the GWAS meta-analysis approach, the first to our knowledge for PiB-PET.

The *APOE* region showed the most significant association where several SNPs surpassed the genome-wide significant threshold ($P < 5E-08$) with *APOE*4* as the top hit that was associated with higher PiB retention in the brain (P -meta=9.09E-30; β =0.18). *APOE*2*, a protective genetic factor against AD, was associated with lower PiB retention, albeit, not genome-wide significant (P -meta=6.57E-05; β =-0.09). This observation is consistent with earlier reports of the association of the *APOE*2/3/4* polymorphism with A β deposition in the brain as measured by PiB-PET¹⁷⁻¹⁹ or florbetapir-PET.³² Likewise, a GWAS of cerebrospinal fluid (CSF) A β has identified a genome-wide significant SNP that was a proxy for *APOE*4*.³³ Numerous prior studies have investigated the role of the *APOE*2/3/4* polymorphism on A β production, aggregation and clearance in the brain,³⁴ but recent studies provide solid mechanistic clues into the role of *APOE* genetic variation in affecting APP transcription and A β production,³⁵ and seeding of amyloid pathology.³⁶ In addition to the *APOE*2/3/4* association, conditional analysis on *APOE*4* identified 14 independent signals in the *APOE* region that also affect brain amyloidosis. Nine of 14 SNPs had essentially no LD with *APOE*4* and *APOE*2* and the remaining 5 showed moderate to weak LD with *APOE*4*. Thus, our meta-analysis indicates the presence of additional signals in the *APOE* region, beyond the *APOE*4*/rs429358 and *APOE*2*/rs7412 SNPs, that affect A β deposition in the brain.

Outside the *APOE* region, the meta-analysis revealed 15 suggestive non-*APOE* loci with $P < 1E-05$ on nine chromosomes. Although they do not meet the established genome-wide significance criteria, their consistent and directional associations in three independent datasets (Table 1) suggest that at least some of them are likely candidate loci for brain amyloidosis process and/or AD risk and variants in these loci may have achieved the genome-wide significance threshold in larger datasets. Credence to this idea was provided by our observation that most of these suggestive loci were also associated with AD risk when we examined the A β -associated SNPs in a published AD GWAS²⁹ (Supplementary Tables S4.1-S4.2). The most significant non-*APOE* SNP (rs13260032; P =4.87E-07) on chromosome 8 is intergenic, and this was an eQTL for a nearby *ADCY8* gene in frontal cortex, which is one of the highest PiB uptake cortical regions.³⁰ *ADCY8* is essential to long-term potentiation and synaptic plasticity and is implicated in memory and learning.³⁷ Genetic variation in or around *ADCY8* has shown to be associated with dissociation symptoms in subjects with posttraumatic stress disorder,³⁷ abdominal visceral³⁸ and alcohol-dependent depression.³⁹ The second top SNP (rs4680057; P =9.69E-07) resides near *C3orf79* and was an eQTL for a nearby long noncoding RNA (lncRNA) gene in anterior cingulate cortex and hippocampus in the brain and for *ARHGEF26* in blood. lncRNAs play a critical role in gene regulatory networks and may affect diverse biological processes and diseases,⁴⁰ including AD where several lncRNAs have been shown to regulate A β production/generation.^{41, 42} A recent GWAS has identified *ARHGEF26* as a new genetic factor for coronary artery disease risk that influences the transendothelial migration of leukocytes.⁴³ The third top SNP (rs12908891; P =1.39E-06) is located *DAPK2* on chromosome 15 that belongs to a family of related serine/threonine kinases shown to be involved in multiple functions, including apoptosis, autophagy, tumor suppression and inflammation.⁴⁴ Although

the role of *DAPK2* in amyloidosis is unknown, another family member, *DAPK1*, promotes the phosphorylation and amyloidogenic processing of APP.⁴⁵ The *DAPK2* region contains other candidate genes, such as *GSNK1G1* and *TRIP4*. While *TRIP4* is a known gene for AD,⁴⁶ *GSNK1G1* has been implicated in the formation of A β .⁴⁷ The top SNP was the most significant eQTL for *HERC1* gene expression in anterior cingulate cortex ($P=7.02E-05$; $P_{SMR}=1.94E-03$). *HERC1* belongs to the ubiquitin–proteasome system that plays a key role in the protein degradation pathway essential for neuronal homeostasis, synaptic development and maintenance. Mutations in *HERC1* have been associated with intellectual disability⁴⁸ and autism spectrum disorders.⁴⁹

To identify additional PiB-relevant candidate genes, we combined results from the brain expression, differential brain expression in AD, eQTL/SMR in the brain, and pathway analyses. Four genes meeting all these functional criteria were identified: *RPS27L* in the *DAPK2* region, *CYP4V2* and *TLR3* in the *CYP4V2* region, and *IDH1* in the *IDH1/C2orf80* region (Table 3, Supplementary Table S6). *RPS27L* is an evolutionarily conserved ribosomal protein and a physiological regulator of transcription factor p53 that is involved in genomic stability and tumor suppression.⁵⁰ p53 has also been implicated in AD progression, in part, due to its interaction with A β in AD progression.⁵¹ p53 also interact with *IDH1* in glioblastoma.⁵² It seems that the involvement of *RPS27L* and *IDH1* in the amyloidogenic process is through their effect on or interaction with p53. Although the role of *CYP4V2* in amyloidosis is currently unclear, activated *TLR3*, along with some members of the toll-like receptors family, can induce A β uptake or inflammatory response during the AD progression.⁵³ Further functional characterization of these candidate genes may help to elucidate their roles in brain amyloidosis.

A recent GWAS using CSFA β 42 as an endophenotype has identified two novel loci in addition to the *APOE* locus.³³ One locus is near *GLIS1* on chromosome 1 and the other in *SERPINB1* on chromosome 6. The reported *GLIS1/185031519* SNP was neither present in our genotyping array nor was it imputed. This SNP was also not in high LD with other SNPs. On the other hand, the reported *SERPINB1/rs316341* SNP was present in our data, but it was not significant ($P=0.148$). We also examined four additional reported *SERPINB1* SNPs with $P<1E-05$ (rs316339, rs316337, rs392120, rsrs2293772)³³ and found one of them to be nominally significant in our data (rs392120; $P=0.033$).

We estimated the genetic variance of global PiB retention explained by the *APOE* and top 15 non-*APOE* SNPs with $P<1E-05$ using a linear regression model. The non-*APOE* SNPs along with *APOE*4* explained 25%–35% of the amyloid variance; of which 14–17% was explained by *APOE*4* alone. A previous study using a different amyloid tracer (florbetapir-PET)³² found a similar contribution of *APOE*4* (11%) to amyloid variance. However, a GWAS on CSF A β 42 found a smaller contribution of *APOE*4* (4%) to amyloid variance.³³ This may be due to the use of different methods to estimate the amyloid variance. While the CSF study used the Genome-wide Complex Trait Analysis (GCTA) that requires >3,000 sample size,⁵⁴ the two amyloid tracer studies with smaller sample sizes used linear regression. Our data, in conjunction with previous studies, highlight the presence of yet to be discovered variants that may be responsible for the unexplained genetic variance of amyloid deposition.

As with any genome-wide study, this study has limitations. Although the present study used the largest combined sample of PiB-PET imaging data reported to-date (from three different centers and ADNI), the sample size was relatively small to achieve genome-wide significance for loci with small effect sizes. We predict that at least some of our suggestive loci with $P < 1E-05$ might have achieved genome-wide significance with a larger sample size, as the direction of allelic effects for all suggestive loci were consistent in all datasets. Unlike some other phenotypes where data could be obtained readily on large numbers of subjects at a relatively low-cost, this is not the case with amyloid PET. Thus, the lack of a very large PiB-PET imaging database for a genome-wide study was a significant constraint. As more PiB-PET imaging data are obtained by different centers, future collaborative studies, as done here, on larger samples may allow the identification of additional genes for brain amyloidosis.

In conclusion, this is the first GWAS on PiB-PET that has confirmed the established association of the *APOE* locus with *in vivo* brain amyloidosis. In addition to the known association, we have identified novel variants in the *APOE* region that affect amyloidosis. A combination of genetic and functional approaches has also led to the identification of additional putative candidate genes that warrant follow-up genetic and functional studies to confirm their role in brain amyloidosis.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

ACKNOWLEDGMENTS

This study was supported by the US National Institutes of Health grants AG030653, AG041718, AG005133, AG025516, AG025204, AG044546, AG003991, AG053303, AG054936, LM012535, AG05681, G026276, and AG024904 and DOD ADNI (Department of Defense award W81XWH-12-2-0012). See full acknowledgments in Supplementary Material

REFERENCES

1. Naj AC, Jun G, Beecham GW, Wang LS, Vardarajan BN, Buross J et al. Common variants at MS4A4/MS4A6E, CD2AP, CD33 and EPHA1 are associated with late-onset Alzheimer's disease. *Nat Genet* 2011; 43(5): 436–441. [PubMed: 21460841]
2. Lambert JC, Ibrahim-Verbaas CA, Harold D, Naj AC, Sims R, Bellenguez C et al. Meta-analysis of 74,046 individuals identifies 11 new susceptibility loci for Alzheimer's disease. *Nat Genet* 2013; 45(12): 1452–1458. [PubMed: 24162737]
3. Ridge PG, Hoyt KB, Boehme K, Mukherjee S, Crane PK, Haines JL et al. Assessment of the genetic variance of late-onset Alzheimer's disease. *Neurobiol Aging* 2016; 41: 200 e213–200 e220.:
4. Hyman BT, Phelps CH, Beach TG, Bigio EH, Cairns NJ, Carrillo MC et al. National Institute on Aging-Alzheimer's Association guidelines for the neuropathologic assessment of Alzheimer's disease. *Alzheimers Dement* 2012; 8(1): 1–13. [PubMed: 22265587]
5. Jack CR, Jr, Knopman DS, Jagust WJ, Petersen RC, Weiner MW, Aisen PS et al. Tracking pathophysiological processes in Alzheimer's disease: an updated hypothetical model of dynamic biomarkers. *Lancet Neurol* 2013; 12(2): 207–216. [PubMed: 23332364]
6. Jack CR, Jr, Therneau TM, Wiste HJ, Weigand SD, Knopman DS, Lowe VJ et al. Transition rates between amyloid and neurodegeneration biomarker states and to dementia: a population-based, longitudinal cohort study. *Lancet Neurol* 2016; 15(1): 56–64. [PubMed: 26597325]

7. Burnham SC, Bourgeat P, Dore V, Savage G, Brown B, Laws S et al. Clinical and cognitive trajectories in cognitively healthy elderly individuals with suspected non-Alzheimer's disease pathophysiology (SNAP) or Alzheimer's disease pathology: a longitudinal study. *Lancet Neurol* 2016; 15(10): 1044–1053. [PubMed: 27450471]
8. Klunk WE, Engler H, Nordberg A, Wang Y, Blomqvist G, Holt DP et al. Imaging brain amyloid in Alzheimer's disease with Pittsburgh Compound-B. *Ann Neurol* 2004; 55(3): 306–319. [PubMed: 14991808]
9. Price JC, Klunk WE, Lopresti BJ, Lu X, Hoge JA, Ziolkowski SK et al. Kinetic modeling of amyloid binding in humans using PET imaging and Pittsburgh Compound-B. *J Cereb Blood Flow Metab* 2005; 25(11): 1528–1547. [PubMed: 15944649]
10. Jack CR, Jr, Barrio JR, Kepe V. Cerebral amyloid PET imaging in Alzheimer's disease. *Acta Neuropathol* 2013; 126(5): 643–657. [PubMed: 24100688]
11. Clark CM, Pontecorvo MJ, Beach TG, Bedell BJ, Coleman RE, Doraiswamy PM et al. Cerebral PET with florbetapir compared with neuropathology at autopsy for detection of neuritic amyloid-beta plaques: a prospective cohort study. *Lancet Neurol* 2012; 11(8): 669–678. [PubMed: 22749065]
12. Sabri O, Sabbagh MN, Seibyl J, Barthel H, Akatsu H, Ouchi Y et al. Florbetaben PET imaging to detect amyloid beta plaques in Alzheimer's disease: phase 3 study. *Alzheimers Dement* 2015; 11(8): 964–974. [PubMed: 25824567]
13. Curtis C, Gamez JE, Singh U, Sadowsky CH, Villena T, Sabbagh MN et al. Phase 3 trial of flutemetamol labeled with radioactive fluorine 18 imaging and neuritic plaque density. *JAMA Neurol* 2015; 72(3): 287–294. [PubMed: 25622185]
14. Blennow K, Mattsson N, Scholl M, Hansson O, Zetterberg H. Amyloid biomarkers in Alzheimer's disease. *Trends Pharmacol Sci* 2015; 36(5): 297–309. [PubMed: 25840462]
15. Kamboh MI, Barmada MM, Demirci FY, Minster RL, Carrasquillo MM, Pankratz VS et al. Genome-wide association analysis of age-at-onset in Alzheimer's disease. *Mol Psychiatry* 2012; 17(12): 1340–1346. [PubMed: 22005931]
16. Naj AC, Jun G, Reitz C, Kunkle BW, Perry W, Park YS et al. Effects of multiple genetic loci on age at onset in late-onset Alzheimer disease: a genome-wide association study. *JAMA Neurol* 2014; 71(11): 1394–1404. [PubMed: 25199842]
17. Mathis CA, Kuller LH, Klunk WE, Snitz BE, Price JC, Weissfeld LA et al. In vivo assessment of amyloid-beta deposition in nondemented very elderly subjects. *Ann Neurol* 2013; 73(6): 751–761. [PubMed: 23596051]
18. Nebes RD, Snitz BE, Cohen AD, Aizenstein HJ, Saxton JA, Halligan EM et al. Cognitive aging in persons with minimal amyloid-beta and white matter hyperintensities. *Neuropsychologia* 2013; 51(11): 2202–2209. [PubMed: 23911776]
19. Morris JC, Roe CM, Xiong C, Fagan AM, Goate AM, Holtzman DM et al. APOE predicts amyloid-beta but not tau Alzheimer pathology in cognitively normal aging. *Ann Neurol* 2010; 67(1): 122–131. [PubMed: 20186853]
20. Swaminathan S, Shen L, Risacher SL, Yoder KK, West JD, Kim S et al. Amyloid pathway-based candidate gene analysis of [(11)C]PiB-PET in the Alzheimer's Disease Neuroimaging Initiative (ADNI) cohort. *Brain Imaging Behav* 2012; 6(1): 1–15. [PubMed: 21901424]
21. Saykin AJ, Shen L, Yao X, Kim S, Nho K, Risacher SL et al. Genetic studies of quantitative MCI and AD phenotypes in ADNI: Progress, opportunities, and plans. *Alzheimers Dement* 2015; 11(7): 792–814. [PubMed: 26194313]
22. Jagust WJ, Bandy D, Chen K, Foster NL, Landau SM, Mathis CA et al. The Alzheimer's Disease Neuroimaging Initiative positron emission tomography core. *Alzheimers Dement* 2010; 6(3): 221–229. [PubMed: 20451870]
23. Deters KD, Risacher SL, Yoder KK, Oblak AL, Unverzagt FW, Murrell JR et al. [(11)C]PiB PET in Gerstmann-Straussler-Scheinker disease. *Am J Nucl Med Mol Imaging* 2016; 6(1): 84–93. [PubMed: 27069768]
24. Risacher SL, Kim S, Shen L, Nho K, Foroud T, Green RC et al. The role of apolipoprotein E (APOE) genotype in early mild cognitive impairment (E-MCI). *Front Aging Neurosci* 2013; 5: 11. [PubMed: 23554593]

25. Mintun MA, Larossa GN, Sheline YI, Dence CS, Lee SY, Mach RH et al. [11C]PIB in a nondemented population: potential antecedent marker of Alzheimer disease. *Neurology* 2006; 67(3): 446–452. [PubMed: 16894106]
26. Howie BN, Donnelly P, Marchini J. A flexible and accurate genotype imputation method for the next generation of genome-wide association studies. *PLoS Genet* 2009; 5(6): e1000529. [PubMed: 19543373] :
27. Genomes Project C, Abecasis GR, Altshuler D, Auton A, Brooks LD, Durbin RM et al. A map of human genome variation from population-scale sequencing. *Nature* 2010; 467(7319): 1061–1073. [PubMed: 20981092]
28. Willer CJ, Li Y, Abecasis GR. METAL: fast and efficient meta-analysis of genomewide association scans. *Bioinformatics* 2010; 26(17): 2190–2191. [PubMed: 20616382]
29. Kamboh MI, Demirci FY, Wang X, Minster RL, Carrasquillo MM, Pankratz VS et al. Genome-wide association study of Alzheimer's disease. *Transl Psychiatry* 2012; 2: e117. [PubMed: 22832961] :
30. Cohen AD, Mowrey W, Weissfeld LA, Aizenstein HJ, McDade E, Mountz JM et al. Classification of amyloid-positivity in controls: comparison of visual read and quantitative approaches. *Neuroimage* 2013; 71: 207–215. [PubMed: 23353602]
31. de Leeuw CA, Mooij JM, Heskes T, Posthuma D. MAGMA: generalized gene-set analysis of GWAS data. *PLoS Comput Biol* 2015; 11(4): e1004219. [PubMed: 25885710] :
32. Ramanan VK, Risacher SL, Nho K, Kim S, Swaminathan S, Shen L et al. APOE and BCHE as modulators of cerebral amyloid deposition: a florbetapir PET genome-wide association study. *Mol Psychiatry* 2014; 19(3): 351–357. [PubMed: 23419831]
33. Deming Y, Li Z, Kapoor M, Harari O, Del-Aguila JL, Black K et al. Genome-wide association study identifies four novel loci associated with Alzheimer's endophenotypes and disease modifiers. *Acta Neuropathol* 2017; 133(5): 839–856. [PubMed: 28247064]
34. Yu JT, Tan L, Hardy J. Apolipoprotein E in Alzheimer's disease: an update. *Annu Rev Neurosci* 2014; 37: 79–100. [PubMed: 24821312]
35. Huang YA, Zhou B, Wernig M, Sudhof TC. ApoE2, ApoE3, and ApoE4 Differentially Stimulate APP Transcription and Abeta Secretion. *Cell* 2017; 168(3): 427–441 e421. [PubMed: 28111074]
36. Liu CC, Zhao N, Fu Y, Wang N, Linares C, Tsai CW, Bu G. ApoE4 accelerates early seeding of amyloid pathology. *Neuron* 2017; 96:1024–1032 [PubMed: 29216449]
37. Wolf EJ, Rasmusson AM, Mitchell KS, Logue MW, Baldwin CT, Miller MW. A genome-wide association study of clinical symptoms of dissociation in a trauma-exposed sample. *Depress Anxiety* 2014; 31(4): 352–360. [PubMed: 24677629]
38. Sung YJ, Perusse L, Sarzynski MA, Fornage M, Sidney S, Sternfeld B et al. Genome-wide association studies suggest sex-specific loci associated with abdominal and visceral fat. *Int J Obes (Lond)* 2016; 40(4): 662–674. [PubMed: 26480920]
39. Procopio DO, Saba LM, Walter H, Lesch O, Skala K, Schlaff G et al. Genetic markers of comorbid depression and alcoholism in women. *Alcohol Clin Exp Res* 2013; 37(6): 896–904. [PubMed: 23278386]
40. Batista PJ, Chang HY. Long noncoding RNAs: cellular address codes in development and disease. *Cell* 2013; 152(6): 1298–1307. [PubMed: 23498938]
41. Luo Q, Chen Y. Long noncoding RNAs and Alzheimer's disease. *Clin Interv Aging* 2016; 11: 867–872. [PubMed: 27418812]
42. Shi C, Zhang L, Qin C. Long non-coding RNAs in brain development, synaptic biology, and Alzheimer's disease. *Brain Res Bull* 2017; 132: 160–169. [PubMed: 28347717]
43. Klarin D, Zhu QM, Emdin CA, Chaffin M, Horner S, McMillan BJ et al. Genetic analysis in UK Biobank links insulin resistance and transendothelial migration pathways to coronary artery disease. *Nat Genet* 2017; 49(9): 1392–1397. [PubMed: 28714974]
44. Geering B Death-associated protein kinase 2: Regulator of apoptosis, autophagy and inflammation. *Int J Biochem Cell Biol* 2015; 65: 151–154. [PubMed: 26055515]
45. Kim BM, You MH, Chen CH, Suh J, Tanzi RE, Ho Lee T. Inhibition of death-associated protein kinase 1 attenuates the phosphorylation and amyloidogenic processing of amyloid precursor protein. *Hum Mol Genet* 2016; 25(12): 2498–2513. [PubMed: 27094130]

46. Ruiz A, Heilmann S, Becker T, Hernandez I, Wagner H, Thelen M et al. Follow-up of loci from the International Genomics of Alzheimer's Disease Project identifies TRIP4 as a novel susceptibility gene. *Transl Psychiatry* 2014; 4: e358. [PubMed: 24495969]
47. Flajolet M, He G, Heiman M, Lin A, Nairn AC, Greengard P. Regulation of Alzheimer's disease amyloid-beta formation by casein kinase I. *Proc Natl Acad Sci U S A* 2007; 104(10): 4159–4164. [PubMed: 17360493]
48. Utine GE, Taskiran EZ, Kosukcu C, Karaosmanoglu B, Guleray N, Dogan OA et al. HERC1 mutations in idiopathic intellectual disability. *Eur J Med Genet* 2017; 60(5): 279–283. [PubMed: 28323226]
49. Hashimoto R, Nakazawa T, Tsurusaki Y, Yasuda Y, Nagayasu K, Matsumura K et al. Whole-exome sequencing and neurite outgrowth analysis in autism spectrum disorder. *J Hum Genet* 2016; 61(3): 199–206. [PubMed: 26582266]
50. Xiong X, Zhao Y, Tang F, Wei D, Thomas D, Wang X et al. Ribosomal protein S27-like is a physiological regulator of p53 that suppresses genomic instability and tumorigenesis. *Elife* 2014; 3: e02236. [PubMed: 25144937]
51. Jazvinscak Jembrek M, Slade N, Hof PR, Simic G. The interactions of p53 with tau and Ass as potential therapeutic targets for Alzheimer's disease. *Prog Neurobiol* 2018.
52. Chaurasia A, Park SH, Seo JW, Park CK. Immunohistochemical Analysis of ATRX, IDH1 and p53 in Glioblastoma and Their Correlations with Patient Survival. *J Korean Med Sci* 2016; 31(8): 1208–1214. [PubMed: 27478330]
53. Gambuzza ME, Sofo V, Salmeri FM, Soraci L, Marino S, Bramanti P. Toll-like receptors in Alzheimer's disease: a therapeutic perspective. *CNS Neurol Disord Drug Targets* 2014; 13(9): 1542–1558. [PubMed: 25106635]
54. Visscher PM, Hemani G, Vinkhuyzen AA, Chen GB, Lee SH, Wray NR et al. Statistical power to detect genetic (co)variance of complex traits using SNP data in unrelated samples. *PLoS Genet* 2014; 10(4): e1004269. [PubMed: 24721987]

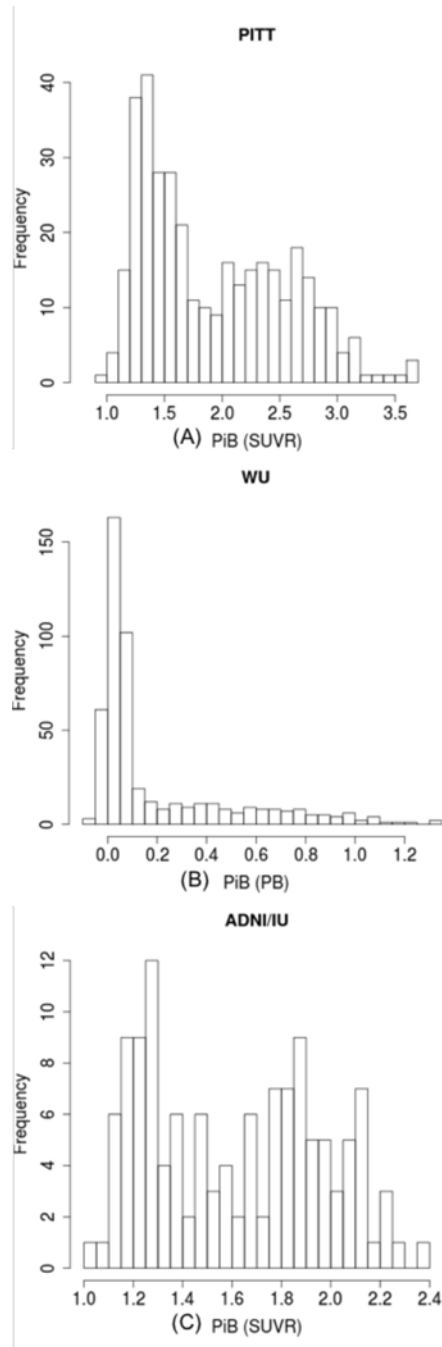


Figure 1. Distribution of PiB retention in the University of Pittsburgh (PITT) (A), Washington University (WU) (B), and the Alzheimer's disease Neuroimaging Initiative (ADNI) and the Indiana Memory and Aging Study (ADNI/IU) (C) samples. SUVR= Standardized Uptake Volume Ratio; BP= Binding Potential

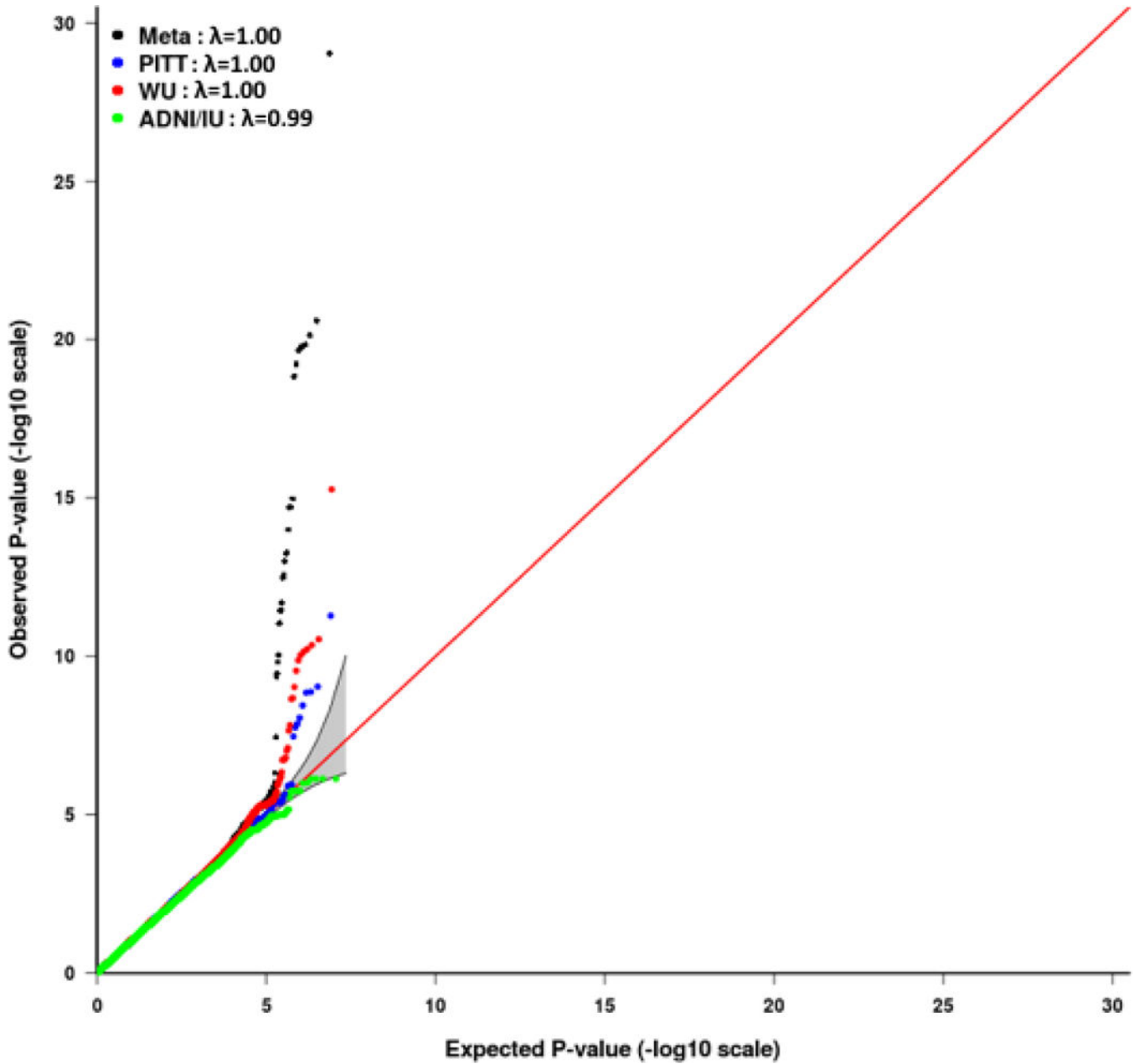


Figure 2a. Quantile-quantile plot for the individual GWAS results in the University of Pittsburgh (PITT), Washington University (WU), and the Alzheimer’s disease Neuroimaging Initiative (ADNI) and the Indiana Memory and Aging Study (ADNI/IU) datasets and in the meta-analysis. λ is the genomic control value.

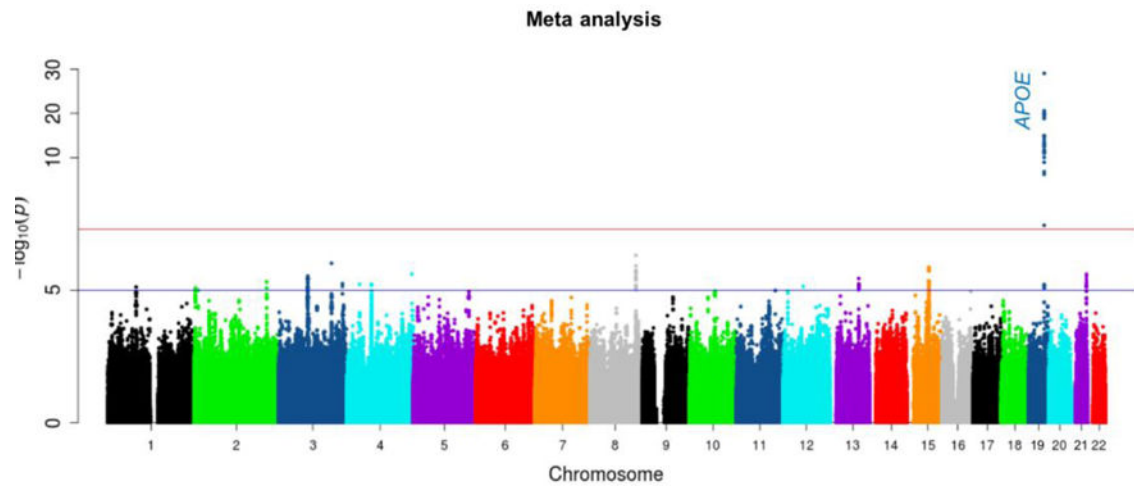


Figure 2b. Manhattan plot showing the P -values in the meta-analysis. The blue line represents the suggestive significance line ($P < E-05$). The red line represents the significance threshold ($P < 5E-08$).

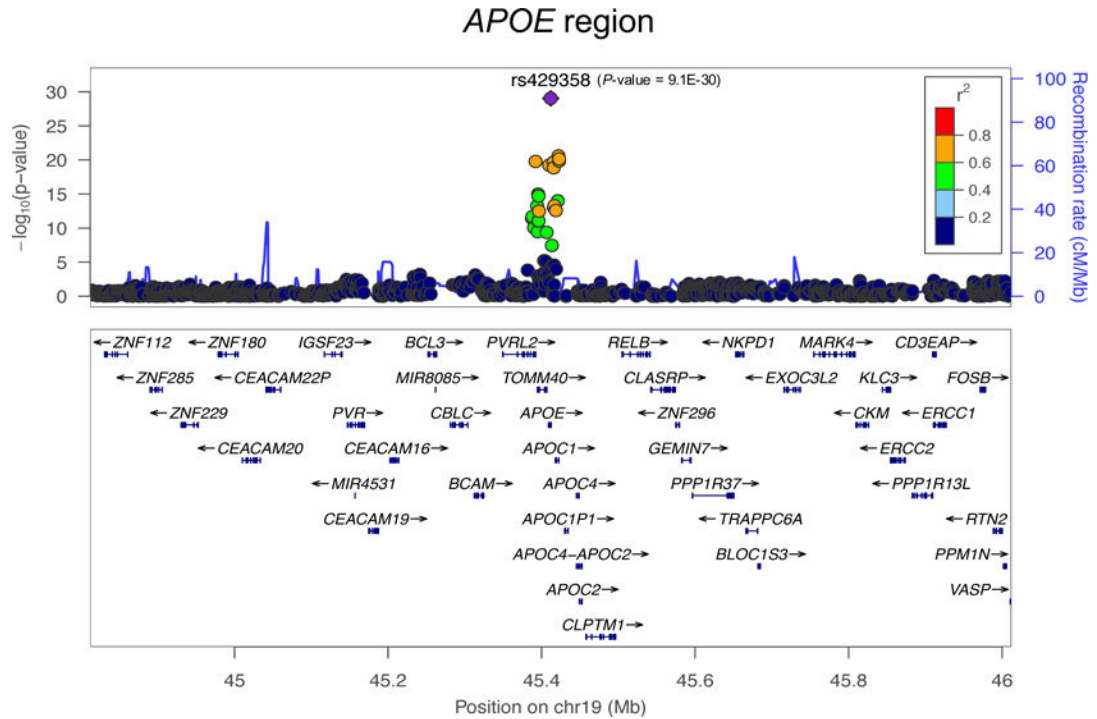


Figure 3. Regional plot of the *APOE* region on chromosome 19 in the meta-analysis. The relative location of genes and the direction of transcription are shown in the lower portion of the figure, and the chromosomal position is shown on the x -axis. The light blue line show the recombination rate across the region (right y -axis), and the left y axis shows the significance of the associations. The purple diamond shows the P -value for rs429358 that is the most significant SNP in the meta-analysis. The circles show the P -values for all other SNPs and are color coded according to the level of LD with rs429358 in the 1000 Genome Project EUR population.

Table 1.

Genetic loci associated with PIB-PET with $P < 1E-05$ in the meta-analysis

SNP	Chr	Position	A1	A2	Gene	Region	PITT			WU			ADN/IU			Meta		Meta adjusted for APOE*4	
							MAF	Beta	P-value	MAF	Beta	P-value	MAF	Beta	P-value	Beta	P-value	Beta	P-value
rs429358	19	45411941	C	T	APOE	exonic	0.20	0.35	5.20E-12	0.21	0.16	5.36E-16	0.28	0.19	7.88E-05	0.18	9.09E-30	NA	NA
rs13260032	8	132451455	C	A	ADCY8, EFR3A	intergenic	0.43	-0.17	4.90E-05	0.44	-0.06	1.34E-02	0.43	-0.07	6.76E-02	-0.08	4.87E-07	-0.07	1.15E-05
rs4680057	1	153096985	A	G	RAP2B, C3orf79	intergenic	0.45	0.16	2.60E-04	0.42	0.04	1.93E-02	0.46	0.11	8.07E-03	0.06	9.69E-07	0.05	2.61E-05
rs12908891	5	64236441	G	A	DAPK2	intronic	0.46	0.12	6.38E-03	0.49	0.06	5.77E-06	0.50	0.01	9.01E-01	0.06	1.39E-06	0.04	5.30E-04
rs7377304	1	187129780	G	T	CYP4V2	intronic	0.42	0.08	8.84E-02	0.46	0.06	5.98E-05	0.47	0.09	2.83E-02	0.06	2.46E-06	0.06	1.59E-05
rs5708341	1	45627581	T	A	C21orf33, ICOSLG	intergenic	0.23	0.18	1.88E-04	0.18	0.05	4.65E-03	0.19	0.07	2.26E-01	0.07	2.51E-06	0.06	6.74E-05
rs9831119	3	84712077	C	T	LINC00971	ncRNA_intronic	0.12	-0.13	3.66E-02	0.14	-0.07	5.15E-04	0.12	-0.16	8.89E-03	-0.08	2.98E-06	-0.07	1.80E-05
rs9531483	3	84244873	A	C	SLITRK1	intergenic	0.30	-0.12	7.81E-03	0.31	-0.06	5.62E-04	0.33	-0.07	1.12E-01	-0.06	3.65E-06	-0.05	8.45E-05
rs6722000	2	209075957	G	A	C2orf80, IDH1	intergenic	0.21	0.14	7.27E-03	0.20	0.07	3.92E-04	0.22	0.06	2.37E-01	0.07	4.96E-06	0.07	1.71E-05
rs11923588	3	184459667	T	C	MAGEF1, LOC101928992	intergenic	0.07	-0.20	2.19E-02	0.06	-0.19	6.50E-05	0.06	-0.14	1.53E-01	-0.18	5.66E-06	-0.13	1.56E-03
rs66837203	7	36897136	T	C	DTHD1, MIR4801	intergenic	0.06	0.24	9.85E-03	0.08	0.10	3.53E-04	0.04	0.13	2.36E-01	0.11	6.03E-06	0.09	4.66E-04
rs200028958	4	70923661	A	G	HTN1	intronic	0.10	0.29	2.99E-05	0.11	0.09	1.29E-03	0.09	-0.05	5.54E-01	0.10	6.25E-06	0.09	8.02E-05
rs4526799	6	57280586	T	C	HSD17B6, SDR9C7	intergenic	0.34	-0.19	3.58E-05	0.34	-0.04	3.09E-02	0.36	-0.04	4.18E-01	-0.05	7.26E-06	-0.06	1.16E-06
rs17105538	1	81315043	G	A	ELTD1, LPHN2	intergenic	0.13	0.13	3.65E-02	0.15	0.08	2.08E-04	0.17	0.09	1.10E-01	0.08	7.66E-06	0.07	1.58E-05
rs62121100	1	3093952	G	T	LINC01250	ncRNA_intronic	0.19	-0.20	1.09E-04	0.16	-0.05	1.03E-02	0.21	-0.04	4.79E-01	-0.07	8.44E-06	-0.06	2.90E-05
rs1809136	2	11152180	G	C	KCNFL1, FLJ33534	intergenic	0.07	0.28	5.91E-04	0.07	0.17	5.45E-04	0.08	0.00	9.95E-01	0.16	9.99E-06	0.15	6.36E-05

A1: minor allele; A2: major allele.

Table 2.

Conditional analysis on SNPs reaching $P < 0.05$ in the *APOE* region with additional adjustment for *APOE**4(rs429358) in the meta-analysis

SNP	Chr	Position	A1	A2	Gene	Region	PITT			WU			ADNI/IU			Meta		LD (R ²) with	
							MAF	Beta	P-value	MAF	Beta	P-value	MAF	Beta	P-value	Beta	P-value	E*4	E*2
rs7412	19	45412079	T	C	<i>APOE</i>	exonic	0.07	-0.21	7.63E-03	0.07	-0.04	1.45E-01	0.04	-0.06	5.49E-01	-0.06	3.69E-03	0.01	NA
rs3852859	19	45379309	C	T	<i>PVRL2</i>	intronic	0.20	0.08	1.14E-01	0.20	0.05	1.10E-01	0.24	0.07	1.52E-01	0.06	8.81E-03	0.004	0.01
rs2075642	19	45377467	A	G	<i>PVRL2</i>	intronic	0.20	0.07	1.78E-01	0.20	0.06	8.44E-02	0.22	0.07	1.33E-01	0.06	1.11E-02	0.004	0.01
rs3852856	19	45361574	A	G	<i>PVRL2</i>	intronic	0.21	0.06	2.43E-01	0.20	0.07	3.16E-02	0.22	0.06	2.03E-01	0.06	1.15E-02	0.006	0.01
rs75627662	19	45413576	T	C	<i>APOE</i>	downstream	0.23	-0.15	1.57E-02	0.23	-0.02	2.92E-01	0.26	-0.03	6.19E-01	-0.03	1.50E-02	0.42	0.21
rs4803767	19	45372959	T	C	<i>PVRL2</i>	intronic	0.26	0.05	2.63E-01	0.28	0.04	1.56E-01	0.31	0.07	8.34E-02	0.05	2.06E-02	0.002	0.01
rs60389450	19	45372184	C	A	<i>PVRL2</i>	intronic	0.26	0.04	3.20E-01	0.26	0.02	1.72E-01	0.31	0.07	8.34E-02	0.03	2.72E-02	0.003	0.01
rs59325138	19	45416291	T	C	<i>APOE,APOC1</i>	intergenic	0.38	0.06	2.16E-01	0.36	0.02	2.50E-01	0.36	0.07	1.17E-01	0.03	3.10E-02	0.15	0.03
rs483082	19	45416178	T	G	<i>APOE,APOC1</i>	intergenic	0.26	-0.10	1.60E-01	0.29	-0.04	1.59E-01	0.32	-0.06	4.51E-01	-0.05	3.35E-02	0.64	0.16
rs8104483	19	45372354	G	T	<i>PVRL2</i>	intronic	0.27	0.03	4.65E-01	0.27	0.02	2.04E-01	0.32	0.09	4.20E-02	0.03	3.67E-02	0.003	0.01
rs3729640	19	45381917	T	C	<i>PVRL2</i>	UTR3	0.20	0.08	9.33E-02	0.19	0.03	3.35E-01	0.23	0.04	4.33E-01	0.05	3.79E-02	0.004	0.01
rs8104292	19	45372707	A	G	<i>PVRL2</i>	intronic	0.27	0.04	4.14E-01	0.28	0.03	2.62E-01	0.32	0.09	4.20E-02	0.04	3.80E-02	0.003	0.01
rs438811	19	45416741	T	C	<i>APOE,APOC1</i>	intergenic	0.26	-0.10	1.60E-01	0.30	-0.04	1.84E-01	0.32	-0.06	4.51E-01	-0.04	3.84E-02	0.65	0.16
rs58521715	19	45372129	T	A	<i>PVRL2</i>	intronic	0.27	0.03	4.65E-01	0.27	0.02	2.53E-01	0.32	0.09	4.20E-02	0.03	4.50E-02	0.004	0.01

A1: minor allele; A2: major allele.

Table 3.

The functional analysis results for target genes.

Out of 257 target genes, only genes meeting at least three functional criteria are listed. The criteria include: 1) differential expression in at least two Alzheimer disease studies, and up- or down-regulated consistently in different studies; 2) expression in brain cells (Barres website); 3) having *cis*-eQTL effects in any brain tissues using GTEx database ($P < 0.05$); 4) mediating genetic effects on PiB (SMR analysis with $P < 0.05$) in any brain tissues; 5) having *cis*-eQTL effects in whole blood ($P < 0.05$); 6) mediating genetic effects on PiB (SMR analysis with $P < 0.05$) in whole blood; and 7) included in nominally significant pathways. The detailed results are summarized in Supplementary Table S6.

Locus	Target gene	Chr	Expression in brain (Barres)	Differential expression in AD	eQTL in blood	SMR in blood	eQTL in any brain tissues	SMR in any brain tissues	Pathway	Non-APOE locus pathway
<i>APOE</i>	<i>APOC1</i>	19								
	<i>APOC2</i>	19								
	<i>APOE</i>	19								
	<i>BCAM</i>	19								
	<i>BCL3</i>	19								
	<i>BLOCS3</i>	19								
	<i>CD3EAP</i>	19								
	<i>CEACAM19</i>	19								
	<i>CKM</i>	19								
	<i>CLASRP</i>	19								
	<i>CLPTM1</i>	19								
	<i>ERCC1</i>	19								
	<i>ERCC2</i>	19								
	<i>EXOC3L2</i>	19								
	<i>GEMIN7</i>	19								
	<i>KLC3</i>	19								
	<i>NKPD1</i>	19								
	<i>PPP1R13L</i>	19								
	<i>PVRL2</i>	19								
	<i>TOMM40</i>	19								
	<i>TRAPPC6A</i>	19								

Locus	Target gene	Chr	Expression in brain (Barres)	Differential expression in AD	eQTL in blood	SMR in blood	eQTL in any brain tissues	SMR in any brain tissues	Pathway	Non-APOE locus pathway
	<i>ZNF180</i>	19	Yellow				Yellow	Yellow		
<i>ADCY8, EFR3A</i>	<i>ADCY8</i>	8								
	<i>EFR3A</i>	8								
<i>RAP2B, C3orf179</i>	<i>RAP2B</i>	3								
	<i>DHX36</i>	3		Green	Blue					
<i>DAPK2</i>	<i>CSNK1G1</i>	15								
	<i>DAPK2</i>	15			Blue					
	<i>FBXL22</i>	15			Blue					
	<i>HERC1</i>	15								
	<i>KIAA0101</i>	15			Blue					
	<i>PIIB</i>	15								
	<i>SNX1</i>	15								
	<i>SNX22</i>	15		Green						
	<i>TRIP4</i>	15			Blue					
	<i>USP3</i>	15		Green						
	<i>LACTB</i>	15			Blue					
	<i>RPS27L</i>	15		Green						
	<i>RAB8B</i>	15								
	<i>ZNF609</i>	15		Green						
	<i>PLEKHO2</i>	15								
	<i>ANKDD1A</i>	15		Green						
<i>CYP4V2</i>	<i>CYP4V2</i>	4		Green	Blue					
	<i>FAM149A</i>	4		Green	Blue					
	<i>FAT1</i>	4		Green						
	<i>KLKB1</i>	4			Blue					
	<i>MTNR1A</i>	4								
	<i>SORBS2</i>	4								
	<i>TLR3</i>	4		Green	Blue					

Locus	Target gene	Chr	Expression in brain (Barres)	Differential expression in AD	eQTL in blood	SMR in blood	eQTL in any brain tissues	SMR in any brain tissues	Pathway	Non-APOE locus pathway
<i>C21orf33</i>	<i>CSTB</i>	21	Yellow		Blue		Yellow	Yellow		
<i>ICOSLG</i>	<i>ICOSLG</i>	21	Yellow							
	<i>PFKL</i>	21	Yellow							
	<i>RRP1</i>	21	Yellow							
	<i>TRAPPC10</i>	21	Yellow							
	<i>HSF2BP</i>	21	Yellow		Blue					
	<i>RRP1B</i>	21	Yellow		Blue					
	<i>UBE2G2</i>	21	Yellow							
<i>LINC00971</i>	<i>CADM2</i>	3	Yellow	Green						
<i>SLITRK1</i>	<i>SLITRK1</i>	13	Yellow							
<i>C2orf80, IDH1</i>	<i>C2orf80</i>	2	Yellow	Green						
	<i>CRYGD</i>	2	Yellow							
	<i>FZD5</i>	2	Yellow	Green						
	<i>IDH1</i>	2	Yellow	Green						
	<i>PIKFYVE</i>	2	Yellow		Blue	Blue				
	<i>PLEKHM3</i>	2	Yellow							
	<i>PTH2R</i>	2	Yellow	Green						
<i>MAGEF1</i>	<i>CAMK2N2</i>	3	Yellow							
<i>LOC101928992</i>	<i>CHRD</i>	3			Blue	Blue				
	<i>ECE2</i>	3	Yellow							
	<i>EHHADH</i>	3	Yellow							
	<i>EIF4G1</i>	3	Yellow							
	<i>EPHB3</i>	3	Yellow	Green	Blue					
	<i>FAM131A</i>	3	Yellow							
	<i>VPS8</i>	3	Yellow							
	<i>PARL</i>	3	Yellow		Blue					
	<i>ABCC5</i>	3	Yellow	Green						
<i>HTN1</i>	<i>ARAP2</i>	4	Yellow	Green						

Locus	Target gene	Chr	Expression in brain (Barres)	Differential expression in AD	eQTL in blood	SMR in blood	eQTL in any brain tissues	SMR in any brain tissues	Pathway	Non-APOE locus pathway
	<i>DTHD1</i>	4	Yellow				Yellow	Yellow		
	<i>SULT1B1</i>	4		Green			Yellow	Yellow		
	<i>UTP3</i>	4	Yellow	Green			Yellow	Yellow		
	<i>DCK</i>	4	Yellow	Green			Yellow	Yellow		
	<i>MOB1B</i>	4	Yellow		Blue	Blue	Yellow	Yellow		
<i>HSD17B6</i>	<i>AIP5B</i>	12	Yellow				Yellow	Yellow		
<i>SDR9C7</i>	<i>BAZ2A</i>	12	Yellow	Green			Yellow	Yellow		
	<i>GLS2</i>	12	Yellow		Blue	Blue	Yellow	Yellow		
	<i>HSD17B6</i>	12	Yellow				Yellow	Yellow		
	<i>MIP</i>	12					Yellow	Yellow		
	<i>MYO1A</i>	12					Yellow	Yellow		
	<i>NAB2</i>	12	Yellow		Blue	Blue	Yellow	Yellow		
	<i>NACA</i>	12	Yellow				Yellow	Yellow		
	<i>NDUFA4L2</i>	12	Yellow				Yellow	Yellow		
	<i>PRIMI</i>	12	Yellow				Yellow	Yellow		
	<i>PTGES3</i>	12	Yellow				Yellow	Yellow		
	<i>R3HDM2</i>	12	Yellow		Blue	Blue	Yellow	Yellow		
	<i>RDH16</i>	12			Blue	Blue	Yellow	Yellow		
	<i>SDR9C7</i>	12					Yellow	Yellow		
	<i>SHMT2</i>	12	Yellow	Green			Yellow	Yellow		
	<i>SPRYD4</i>	12	Yellow	Green			Yellow	Yellow		
	<i>STAT6</i>	12	Yellow				Yellow	Yellow		
	<i>TAC3</i>	12	Yellow	Green			Yellow	Yellow		
	<i>TIMELESS</i>	12	Yellow				Yellow	Yellow		
	<i>TMEM194A</i>	12	Yellow				Yellow	Yellow		
	<i>ZBTB39</i>	12	Yellow		Blue	Blue	Yellow	Yellow		
<i>LPHN2</i>	<i>LPHN2</i>	1	Yellow				Yellow	Yellow		
<i>LINC01250</i>	<i>ADII</i>	2	Yellow				Yellow	Yellow		

Locus	Target gene	Chr	Expression in brain (Barres)	Differential expression in AD	eQTL in blood	SMR in blood	eQTL in any brain tissues	SMR in any brain tissues	Pathway	Non-APOE locus pathway
	<i>RNASEH1</i>	2	Yellow	Green			Yellow	Yellow		
	<i>TRAPPPC12</i>	2	Yellow	Green			Yellow	Yellow		
	<i>TSSC1</i>	2	Yellow	Green			Yellow			
<i>KCNF1</i>	<i>ATP6V1C2</i>	2	Yellow				Yellow	Yellow		
<i>FLJ135534</i>	<i>C2orf50</i>	2	Yellow				Yellow	Yellow		
	<i>E2F6</i>	2	Yellow				Yellow	Yellow		
	<i>KCNF1</i>	2	White	Green			Yellow	White	Orange	Orange
	<i>NOL10</i>	2	Yellow				Yellow	Yellow		
	<i>ROCK2</i>	2	Yellow				Yellow	Yellow	Orange	Orange
	<i>GREB1</i>	2	Yellow				Yellow	Yellow		